## **33. The Synthesis of Boronolide**

by **Charles W. Jefford\*** and **Marie-Claude Moulin** 

Department of Organic Chemistry, University of Geneva, CH-1211 Geneva 4

## **(3.** XII. *90)*

Racemic boronolide **(1)** is prepared in six steps in 4.4% overall yield from acrolein dimer **6** and I-(trimethylsily1)hex- I-yne **(8).** The latter, by hydromagnesiation, is condensed with *6* to give the corresponding fhreo-allylic alcohol **13** (Scheme 2). Conversion of **13** to the erythro-allylic alcohol *5* (Scheme *3),* bis-hydroxylation, and acetylation afford **1.** 

**Introduction.** - *Tetradenia fruticosa* is a shrub belonging to the Lamiacea family which grows in Madagascar where it is used for medicinal purposes. In 1971, the active principle was extracted from the bark and branches and identified as boronolide [l]. Fourteen years later the relative configuration of boronolide was determined by X-ray analysis, and the  $(R)$ -configuration was assigned to the  $C(6)$  position by application of *Hudson's* lactone rule to the molecular rotation **[2].** Subsequently, in 1987, by means of chemical degradation, the absolute configuration of boronolide was established as **(l'R,2'R,3'S,6R)-5,6-dihydr0-6-[ 1',2',3'-tris(acetoxy)heptyl]-2H-pyran-2-one (1) [3].** 



Boronolide has also been isolated from the dried leaves of *T. burberae* **[3]** while deacetylated **(2)** and dideacetylated **(3)** derivatives have been obtained from *T. riparia* **[4]**  (formerly *Iboza),* a central-African species widely used as a tribal medicine. Typically, the Zulu employ extracts of the root as an emetic, while an infusion of the leaf has been reported to be effective against malaria [S].

So far no synthesis of **1** in either its racemic or enatiomerically pure form has been reported. On the basis of precedent *[6-8],* acrolein dimer **6** would be an ideal starting point for **1** as the disconnective analysis shows *(Scheme 1).* The triacetate **1,** through its corresponding triol, has as a possible precursor the *erythro* -allylic alcohol **4** from which it would have to be derived by stereoselective bis-hydroxylation. The  $\alpha$ , $\beta$ -unsaturated  $\delta$ -lactone function in **4** could be introduced by appropriate oxidation of the dihydro-2Hpyran 5. Attachment of the side chain should be realizable by addition of the  $(Z)$ -hex-1-



enyl anion **(7)** to acrolein dimer **6,** provided the diastereoselectivity can be controlled to afford the desired erythro -alcohol.

**Results and Discussion.** – The logical choice of reagent for the hexenyl carbanion synthon **7** is the silylated hexenylmagnesium bromide **9.** Previous work [9-111 has shown that Crignard reagents such as **9** by conversion in situ to their copper analogues **10**  undergo stereoselective addition to aldehydes generating  $(E)$ -allylic alcohols. Accordingly, **1-(trimethylsily1)hex-1-yne (8)** [12] was treated with i-BuMgBr in the presence of a catalytic amount of Cp,TiCl,  $(Cp = cyclopentadienyl)$  resulting in its hydromagnesiation to give exclusively the (E)-derivative **9** [13] [I41 (Scheme 2). When **9** was allowed to react



with acrolein dimer 6 in THF at  $-78^{\circ}$  together with a slight excess of CuBr and Me<sub>5</sub>S, addition of the cuprous intermediate **10** occurred with high diastereoselectivity furnishing the threo - and erythro- alcohols **11/12** of (Z)-configuration in a 15 : 1 ratio and in a yield of 66 *YO.* After chromatography, **11** was treated with Bu,NF which effected desilylation giving the threo-alcohol **13** of (E)-configuration in 99% yield [15]. Alcohol **13** was easily distinguishable from its epimer **5** (see below) thanks to their characteristic 3J(H,H) values in the 'H-NMR spectra [16]. The values of 7.5 and 3.5 **Hz** are consistent with the arrangement of the pair of methine protons in the preferred staggered conformations adopted by the threo- and erythro-epimers **13** and *5,* respectively (Fig. I). stic <sup>3</sup> $J(H,H)$  value<br>
onsistent with the<br>
red conformation<br>
1).<br>
Bu





**Fig.** 2. *Chelation-controlled formation of* **11** *from 6* 

Apart from the retention of the geometry of the vinyl entity, the high diastereoselectivity  $(15:1)$  in favor of the *threo*-alcohol 11 is noteworthy and is explicable in terms of a transition state resembling the threo-product. Chelation by Mg or Cu brings the aldehydic and pyran 0-atoms close together, while the vinylcopper reagent 10 attacks the least hindered face of the aldehyde group *(Fig. 2)*. Such stereoselectivity is characteristic of acrolein dimer **[7]** [8] and has also been observed for structurally similar aldehydes such as 2,3-0 **-isopropylideneglyceraldehyde** [9] and 2,3-0 -dibenzylglyceraldehyde [ 171.

Having secured threo-alcohol 13, it needs to be converted to its erythro-epimer 5. The serviceable method of *Mitsunobu* was selected [ 181. Treatment of threo-alcohol 13 with PPh<sub>1</sub>, diethyl diazoacetate, and benzoic acid led to a 1:1 mixture of the allylic *erythro*benzoates 15 and **16** in an overall yield of 71 *YO* (Scheme *3).* The erythro -configuration of 15 was compatible with its <sup>1</sup>H-NMR spectrum. In any event, the  $S<sub>N</sub>2$  reaction of the phosphonium-oxide intermediate 14 with benzoate must occur with inversion. However, in the alternative  $S_N^2$  reaction, benzoate could attack 14, regardless of its preferred conformation, on either the *si* or re faces of the vinyl terminus (Scheme 3). Experimentally, only a single benzoate, designated as 16, was obtained. The correctness of this assignment was later confirmed by its conversion after saponification to 1. The benzoate mixture 15/16 was saponified to the corresponding alcohols **5/17** in 90% yield.



Trial experiments indicated that the photo-oxygenation of erythro -alcohol **5** was straightforward giving hydroperoxide 18 which could be dehydrated to the unsaturated  $\delta$ -lactone 4 (*Scheme 4*). Unfortunately, 4 was not suitable for hydroxylation with OsO<sub>4</sub> as several decomposition products arose from oxidation of the endocyclic double bond. Consequently, we decided to convert the dihydro- $2H$ -pyran ring into the saturated  $\delta$ -lactone **20**. This step was readily accomplished by the acid-catalyzed addition of H<sub>2</sub>O<sub>2</sub> to 5. Dehydration of the intermediate hydroperoxide 19 gave 20 in 90% yield as a viscous white solid (Scheme *4).* 

At first sight, bis-hydroxylation of **20** in the desired stereoselective sense is unpromising. Whatever predictive model is employed  $[19-21]$ , the expectation is that an allylic alcohol of the (E)-configuration such as **20** would suffer osmylation with modernate stereoselectivity to favor the wrong configuration [22] [23]. However, it is entirely possible that the tetrahydro- $2H$ -pyran substituent might exert a countervailing electronic effect and thereby alter the stereoselectivity. Bis-hydroxylation of **20** was accomplished using catalytic quantities of  $OsO<sub>4</sub>$  regenerated by N-methylmorpholine N-oxide [24] [25]. Two triols **21** and **22** having the desired and undesired configurations, respectively, were obtained in a ratio of 1 :3 (Scheme *4).* Treatment of the mixture with Ac,O in the presence



of pyridine gave the corresponding triacetates **23** and **24** in 20 and 59% yield, respectively. This stereoselective result confirms the influence of the tetrahydro-2H-pyran substituent in substantially diminishing the stereoselectivity without actually reversing it. Nonetheless, the triacetate portion of boronolide is introduced with little trouble. Next, the acetates **23** and **24** were easily separated by chromatography so that the endocyclic double bond could be reestablished. Dehydrogenation was brought about using benzeneseleninic anhydride in chlorobenzene at 130" [26]. The triacetate **23** furnished racemic boronolide **1** in 61 % yield as a colorless solid. Its **I3C-** and 'H-NMR spectra were identical with those of the natural material. For the sake of completeness and for comparative purposes, the isomeric triacetate **24** was similarly oxidized to the isomer **25** of boronolide, easily distinguishable from **1** by its different NMR spectra.

Allylic alcohol **17** which accompanied the formation of *5* is also of preparative utility. It too was oxidized with  $H_2O_2$  to  $\delta$ -lactone 26 (Scheme 5). Bis-hydroxylation of the latter proceeded with the same selectivity as before, delivering triols **21** and **27** in a ratio of **1** : 3, which were acetylated to the triacetates **23** and **28** in the same ratio and in *75%* yield.



Proof of the configuration of **23,** and by extrapolation that which was arbitrarily assigned to 17, was secured by carrying out the  $\alpha, \beta$ -dehydration of the  $\delta$ -lactone ring. Once again, boronolide **(1)** was obtained in **63** % yield.

**Conclusion.** - The preparation of boronolide **(1)** has been achieved from acrolein dimer **6** in essentially six steps in an overall yield of 4.4%. Although not attempted, resolution of the racemic product is feasible by chromatography over cellulose triacetate [27]. Consequently, the simplicity and cheapness of the aforementioned procedure recommends it for making other structurally similar naturally occurring  $\delta$ -lactones.

We wish to thank the *Swiss* National Science Foundation for financial support of this work (grant No. 2.812-0.85).

## Experimental Part

*General.* Solvents were purchased from FIuka AG, Aldrich AG, Merck AG, and Tokyo *Kasei,* dried, and purified before use. TLC: silica gel *60 (F254* Merck) coated plates. Column chromatography: Merck silica gel 60 (230-400 mesh ASTM). Gas-phase chromatography: Hewlett-Packard-HP588A chromatograph; anal. capillary columns packed with dimethylsilicone (12.5 and 25 m long and 0.5 mm internal diameter); N<sub>2</sub> flow of 50 ml/min. M.p. Reichert hot-plate instrument; uncorrected. IR spectra: Perkin-Elmer-618 spectrometer; CHCl<sub>3</sub> solns.; absorptions in cm<sup>-1</sup>.<sup>1</sup>H- and <sup>13</sup>C-NMR spectra: Varian-XL-200 and Bruker-WH-360 instruments; CDCl<sub>3</sub> as solvent; chemical shifts  $(\delta)$  in ppm with reference to tetramethylsilane (TMS); signal intensities are normalized to 1 H; coupling constants *(J)* in Hz. MS: Finnigan-4000 and VG-70-70E spectrometers. Elemental analyses were performed by Dr. H.J. Eder, Service de Microchimie, Institut de Chimie Pharmaceutique, University of Geneva.

*(I* RS, *Z)-1-[(2SR)-3,4-Dihydro-ZH-pyran-2-yl]-2-(trimethylsilyl)hept-2-en-l-ol(ll).* Cp,TiCI, (80 mg, 0.3 mmol) was added to a soln. of i-BuMgBr (10 mmol) in Et<sub>2</sub>O (13 ml) at 0° under Ar. After stirring the soln. for 30 min at 0", **I-(trimethylsily1)hex-I-yne** [I21 **(8;** 1.617 g, 10.5 mmol) was added dropwise while stirring was continued for 6 h at 25°. Next, the solvent was evaporated at 15 Torr; the residue was dissolved in THF (50 ml) at  $-70^{\circ}$  after which CuBr<sup>.</sup>Me<sub>2</sub>S (2.47 g, 12 mmol) was added followed by stirring for 30 min. Acrolein dimer 6 (678 mg, 6.05 mmol) was then added dropwise; stirring was continued for 30 min at  $-78^{\circ}$ , thereafter the temp. was allowed to reach 20–25° overnight. The resulting mixture containing threo/erythro-mixture 11/12 (15:1 according to <sup>1</sup>H-NMR) was filtered and separated by chromatography (hexane/AcOEt 15:1) into 11/12 (1.1 g, 66%) followed by pure 11, pale yellow oil (997 mg, 61%). IR: 3500vs, 3060s, 2960-2850v.s, 1650s, 1610m, 1460s, 1240vs, 1070vs, 980m, 850~3, 760m, 730m. 'H-NMR: 0.18 **(s,** 9 H); 0.9 (t, *J* = 7, 3 H); 1.3 *(m,* 10 H); 2.5 *(d, J* = 2, 1 H); 3.8 *(ddd, <sup>J</sup>*= 10, 8, 2, I H); 4.1 *(d, J* = 8, 1 H); 4.7 *(m,* 1 H); 6.25 (t, *J* = 8, 1 H); 6.4 *(d, J* = 6, 1 H). **MS:** *55,* 73 (IOO), 75, 83, 84, 113, 127, 169, 185, 211, 235, 268 *(M<sup>+</sup>)*. Anal. calc. for C<sub>15</sub>H<sub>28</sub>SiO<sub>2</sub>: C 67.11, H 10.51; found: C 67.00, H 10.62.

*(I RS.E)-l-[(2RS)-3,4-Dihydro-2H-pyran-2-yl]hept-2-en-l-ol(13).* To a soln. of 11 (536 g, 2 mmol) in THF (2 ml) under  $N_2$  at 0°, t-BuOK (224 mg, 2 mmol) and Bu<sub>4</sub>NF (2 mmol/2 ml THF) was added. After allowing to stand for 10 min, the soln. was treated with aq.  $NH<sub>4</sub>Cl$  soln. (2 ml). The soln. was extracted with CH<sub>2</sub>Cl<sub>2</sub>, the extracts dried (MgSO<sub>4</sub>), filtered, and evaporated. The residue was purified by chromatography (hexane/AcOEt 10:1) to give 13 as a pale yellow oil (387 mg, 99%). IR: 3580s, 2900vs, 1650s, 1450m, 1390m, 1220vs, 1070s, 970s.  ${}^{1}H\text{-NMR}:0.84(t, J = 7, 3H); 1.3-2.1(m, 10H); 2.48(s, 1H); 3.64(ddd, J = 10, 7.5, 2.5, 1H); 4.0(t, J = 7.5, 1H);$ 4.7 *(m,* **1** H); 5.44(ddt, *J* = 15.5,7.5, 1.5, **1** H); 5.78 *(dt, J* = 15.5,6.5, **1** H); 6.38 *(m,* 1 H). MS: *55,* 57,73 (loo), 75, 83, 95, 105, 113, 119, 127, 133, 147, 169. Anal. calc. for C<sub>12</sub>H<sub>20</sub>O<sub>2</sub>: C 73.43, H 10.27; found: C 73.23, H 10.39.

*(1* RS, *E)-1-[(2SR)-3,4-Dihydro-2H-pyran-2-yl]hept-2-enyl* Benzoate (15) and *(I* RS)-I- {( E)-2-[(2SR)- 3,4-Dihydro-2H-pyran-2-yl/ethenyl\pentyl Benzoate (16). Benzoic acid (135 mg, 1.1 mmol) and diethyl azodicarboxylate (173  $\mu$ l, 192 mg, 1.1 mmol) were added to **13** (196 mg, 1.0 mmol) in Et<sub>2</sub>O (4 ml) under N<sub>2</sub> [18]. To the resulting mixture, PPh<sub>3</sub> (290 mg, 1.1 mmol) in Et<sub>2</sub>O (4 ml) was added dropwise and then stirred at 20° overnight. The suspension was filtered, the filtrate evaporated, and the residue purified by chromatography (hexane/AcOEt 20:l): 15/16(215mg,71%), 1:l ratio.

15: IR:  $3060-2850v_s$ , 1720vs,1650s, 1450m, 1270vs, 1220vs, 1120s, 1070s. <sup>1</sup>H-NMR: 0.86 (t, *J* = 7, 3 H); 1.3 *(m,* 4 H); 1.7-2.2 *(in,* 6 H); 4.02 *(ddd, J* = 11,4, 2, 1 H); 4.7 *(m,* I H); 5.62 *(m.* 2 **H);** 5.9 *(dtd, J* = 14.5, 7, 1.5, I **H);**  6.4 *(m,* **1** H); 7.44 *(m,* 2 H); 7.56 *(m,* 1 H); 8.08 *(m,* 2 H).

16: 1R: same as that of 15. 'H-NMR: *0.86* (t, *J* = 7,3 H); 1.2-1.4 *(m.* 4 H); 1.6-2.1 *(m,* 6 H); 4.36 *(m.* 1 H); 4.7 *(m.* 1 H); 5.52 (dt, *J* = 7, 6, **1** H); 5.82 *(m.* 2 H); 6.4 *(m,* **1** H); 7.44 *(m,* 2 H); 7.56 *(m.* 1 H); 8.08 *(m.* 2 H).

*(I* RS, *E)-l-[(2SR)-3,4-Dihydro-2H-pyrun-2-y/]hept-2-en-l-ol(5) und* (3RS, E *j-1-[(2SR)-3,4-Dihydro-2Hpyrun-2-yl]hept-/-en-3-ol* **(17).** A mixture of **15/16** (38.4 mg, 0.13 mmol) in EtOH (2 ml), KOH (22 mg, *0.55* mmol), and H<sub>2</sub>O (25  $\mu$ ), 1.4 mmol) was boiled under reflux for 50 min. The soln. was extracted with CH<sub>2</sub>Cl<sub>2</sub>, the extracts dried (MgSO<sub>4</sub>), filtered, and evaporated. Chromatography (CH<sub>2</sub>Cl<sub>2</sub>) gave 5 and 17 each as a colorless oil (11.5 and 11.2 mg, resp. ; 90 % yield).

5: IR:  $3600s$ ,  $3000-2860v$ s,  $1650s$ ,  $1450m$ ,  $1220v$ s,  $1070s$ ,  $970m$ . <sup>1</sup>H-NMR: 0.86 (t,  $J = 7$ , 3 H); 1.2-1.4 (m, 4H); 1.C2.2 *(m,* 6H); **3.8** *(ddd, J* = 11, 3.5, 3.5, **1** H); 4.2 *(m,* 1 H); 4.7 *(m,* **1** H); 5.52 *(ddt,J* = 15.5, 7, 1.5, **1** H); *5.8 (dtd, J* = 15.5, 7, 1, 1 H); 6.4 *(m,* **1** H).

**17:** IR: identical to that of **5.** 'H-NMR: 0.86 (t, *J* = 7, 3 H); 1.2-1.4 *(m,* 4 H); 1.4-1.7 *(m,* 4 **H);** 1.8-2.2 *(m,*  **<sup>3</sup>**H); 4.12 (dt, *J* = 6, 6, **1** H); 4.32 *(m,* **1** H); 4.68 *(m,* **1** H); 5.76 *(m,* 2 H); 6.4 *(m,* 1 H).

(6RSj-5,6-Dihydr0-6-[(1 SR, *E)-I-hydroxyhept-2-eny/]-2H-pyrun-2-one* **(4).** A soln. of **5** (392 mg, 2 mmol) and tetraphenylporphin (13 mg) in toluene (6 ml) was irradiated while dry  $O_2$  was passed for 30 min [6]. After addition of Et<sub>3</sub>N (0.24 ml) and Ac<sub>2</sub>O (0.24 ml), the soln. was stirred overnight and then extracted with Et<sub>2</sub>O. The extracts were washed (H<sub>2</sub>O aq. sat. NaCl soln.), dried (NaSO<sub>4</sub>), and evaporated. Chromatography over *Florisil* (hexane/AcOEt 5:2) gave **4** (270 mg, 64% yield). Colorless oil. IR: 3600s, 3000-295Ovs, 1750vs, 1380s, **1250vs,**  1150m,975m. 'H-NMR: 0.86 *(t. J* = 7, 3 H); 1.34 *(m,* 4 H); 2.06 *(m.* 2 H); 2.32 *(dddd, J* = 18,6,3.5, 1, **1** H); 2.44 (s, **<sup>1</sup>**H); 2.60 *(ddt, J* = 18, 12, 2, **1** H); 4.40 *(dq, J* = **18,** 8, 4, 2 H); 5.46 (ddt, *J* = **15,** 7, 1.5, 1 H); 5.84 (dtd, *J* = **15,** 7, *l.5,lH);6.02(ddd,/=9,3.5,1,1H);6.94(ddd,J=9,6,3,1H).* 

Product **4** was submitted to osmylation according to the procedure described below and gave several unidentified oxidation products.

(6RS)-Tetruhydro-6-[(1 SR, *E)-I-hydroxyhept-2-enyl]-2H-pyrun-2-one* **(20).** A soh. of **5** (645 mg, 3.29 mmol) in THF (10 ml) and 30% aq. H<sub>2</sub>O<sub>2</sub> soln. (1.5 ml, 49 mmol) and conc. H<sub>2</sub>SO<sub>4</sub> (2-3 drops) was stirred at 20° for 36 h. The resulting soln. was poured into a sat.  $(NH_4)_2SO_4$  soln. and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined org. layers were washed with sat. NaHCO<sub>3</sub> and sat. NaCl soln., dried (Na<sub>2</sub>SO<sub>4</sub>), and filtered. The filtrate, after treatment with Et<sub>3</sub>N (400 μl, 2.9 mmol) and Ac<sub>2</sub>O (400 μl, 4.2 mmol), was stirred overnight at 20°. The soln. was extracted with CH<sub>2</sub>Cl<sub>2</sub> and the combined extract washed with 5% aq. HCl, sat. NaHCO<sub>3</sub>, and sat. NaCl soln., dried (Na<sub>2</sub>SO<sub>4</sub>), and filtered. Evaporation and chromatography (hexane/AcOEt 5:1) gave 20 (625 mg, 90%). White viscous solid. IR: 3700-3400vs, 2960-2860vs, 1760vs, 1240s, 1050s, 970m, 910m. <sup>1</sup>H-NMR: 0.84 *(i, J* = 7, **<sup>3</sup>**H); 1.2-1.4 **(m,4H);** 1.6-2.1 *(m,* 6H); 2.3-2.5 *(m.* 2 **H);** 2.5-2.64(m, 1 H); 4.3 *(m,* 2 H); 5.44(ddt, *J* = 15.5,7, **1.5, <sup>1</sup>**H); *5.8(dfd,J* = 15.5,7, 1, **1** H). MS: 55,57(100), 71, **81,95,99,** 105, 113, **151, 156,** 194,212(M+). Anal. calc. for  $C_1$ ,  $H_{20}O_3$ : C 67.89, H 9.50; found: C 67.62, H 9.37.

*(I'* RS,? RS,Y SR.6RS)- *und (I'RS,Z'SR,3'RS,6RS)-Tetrahydro-6-[1',2',3'-tris(acetoxy)heptyl]-2H*pyran-2-ones **(23** and **24).** N-Methylmorpholine N-oxide (2.34~ in t-BuOH; 1 ml, 2.34 mmol), H20 (1.22 ml, *68*  mmol), and acetone (0.45 ml, 6.1 mmol) were mixed with **20** (470 mg, 2.22 mmol). Then, OsO<sub>4</sub> (2.5% in t-BuOH; 71 pl, 5.77 mmol) was added [24] [25]. The mixture was stirred for 8 h, treated with aq. NaHSO, soln. (0.3 m, 2 ml) and filtered over Florisil. The pH was adjusted to 7, acetone evaporated, and t-BuOH removed in vacuo. To the resulting residue of 21/22, pyridine (30 ml, 372 mmol) was directly added. After cooling to 0°, Ac<sub>2</sub>O (6 ml, 63 mmol) was added dropwise. The resulting soln. was stirred for 3 days at 0" and then treated with cold *5%* aq. HCI soln. until the pH reached 7. The soln. was then extracted with  $CH_2Cl_2$ , the extracts washed with aq. sat. NaCl soln., dried (MgSO,), filtered, and evaporated, and the residue purified by chromatography (hexane/AcOEt 3 :2): **23**  (143 mg, 20%) and **24** (418 mg, 59%) as a colorless oil and solid (m.p. 79-81°), resp.

**23:** IR: 3020-2860s, 1740vs, 1370m,1220vs, **11OOvs.** 'H-NMR: 0.88 (t, *J* = 7,3 H); 1.2-2.0 *(m,* 10 H); 2.09 **(s, 3H);2.12(~,3H);2.14(~,3H);2.~2.7(m,2H);4.44(ddd,J=11,6,3, 1** H);5.04(dt,J=7,6, lH);5,24(dd,  $J=6,5,1$  H); 5.38 (dd,  $J=6,5,1$  H). MS: 55, 71, 99, 111, 124, 142 (100), 171, 184, 273, 313, 373 ( $[M+1]^+$ ). Anal. calc. for C<sub>18</sub>H<sub>28</sub>O<sub>8</sub>: C 58.05, H 7.58; found: C 57.77, H 7.80.

**24:** IR: identical to that **of23.** 'H-NMR: 0.84 (t, *J* = 7, 3 H); 1.2-2.0 *(m,* 10 H); 2.03 (2s, 6 H); 2.12 **(s,** 3 H); 142 (100), 159, 171, 184, 201, 231, 273, 313, 373 *([M + 1]<sup>+</sup>*). Anal. calc. for C<sub>18</sub>H<sub>28</sub>O<sub>8</sub>: C 58.05, H 7.58; found: C 58.08, H 7.55. 2.3-2.6 (m, 2 H); 4.5 (dt, J = 16, 3.7, 1 H); 5.16 (ddd, J = 8, 5.5, 2, 1 H); 5.3 (m, 2 H). MS: 55, 71, 84, 99, 112, 124,

*(I'RS,2'RS,3'SR,6RS)-5,6-Dihydro-6-[I",3'-tris(ncetoxy)heptyi]-2~-pyrun-2-one* **(1).** A soln. of **23** (41 mg, 0.11 mmol) and benzeneseleninic anhydride (40 mg, 0.11 mmol) in dry chlorobenzene (3 ml) was heated at 130° for 70 h [26]. After cooling, the solvent was evaporated and the residue purified by chromatography (hexane/ AcOEt 3:2): 1 (25 mg, 61%). Colorless solid. M.p. 79-81° ([1]: 90°). IR: 3020-2860s, 1750vs, 1370s, 1220vs, 1050vs. 'H-NMR: 0.84(t, *J* = 7, 3 H); 1.2-1.3 (m,4H); 1.5-1.6 *(m.* **2H);** 2.03 **(s, 3** H); 2.06(s, 3 H);2.1 *(s,* 3 H); 2.2-2.6(m, 2 *H);* 4.54 *(ddd, J* = 12, 6, 4, 1 H); 5.04 *(dt, J* = 6, 6, 1 H); 5.34 *(m,* 2 H); 6.04 *(ddd, J* = 10, 2.5, 1, 1 *H);* 6.88 *(ddd, J* = 10, 6, 2.5, 1 **H**). <sup>13</sup>C-NMR: 13.78 *(q)*; 20.54 *(q)*; 20.64 *(q)*; 22.30 *(t)*; 25.07 *(t)*; 26.98 *(t)*; 30.21 *(t)*; 70.54 *(d)*; 70.61 *(d);* 71.58 *(d);* 75.09 *(d);* 121.40 *(d);* 144.02 *(d);* 162.36 **(s);** 169.60 (8); 169.81 **(s);** 170.36 **(s).** MS: 55,68, 82, 97, 110, 122, 140 (loo), 159, 171, 182,201,242,273,313,371 *([M* + 1]+). Anal. calc. for C,,H,,O,: **C** 58.37, H 7.08; found: C 58.21, *H* 7.05.

(1' RS.2' SR,3' RS.6 *RS)-5,6-Dihydro-6-[ 1'.2',3'-tris (acetoxy) heptyl]-2H-pyran-2-one* (25). The previous experiment was repeated with 24 to give 25 in the same yield. <sup>1</sup>H-NMR: 0.86  $(t, J = 7, 3 H)$ ; 1.2-1.6  $(m, 6 H)$ ; 2.06  $(2s,$ **6H);2.14(~,3H);2.5(m,2H);4.56(dt,J=11,5,1H);5.16(m,1H);5.3(dd,J=8,3,1H);5.42(dd,J=8,5,**  <sup>1</sup>*H);* 6.02 *(ddd, J* = 10, 2.2, 1.3, 1 H); 6.9 *(ddd, J* = 10, *5.5, 3,* 1 *H).* I3C-NMR: 13.82 *(4);* 20.70 (y); 20.94 *(4);*  22.38 *(t);* 24.45 *(t);* 27.09 *(t);* 30.39 *(t);* 30.86 *(4);* 69.36 *(d);* 70.27 *(d);* 70.84 *(d);* 121.06 *(d);* 144.56 *(d);* 162.72 **(s);**  169.09 **(s);** 170.26 **(s);** 170.60 **(s).** IR, MS: identical to those of 1. Anal. calc. for C,,H2,08: C 58.37, H 7.08; found: **C** 58.18, H 7.12.

 $(1'RS,2'RS,3'SR,6RS)$ - and  $(1'SR,2'SR,3'SR,6RS)$ -Tetrahydro-6-[1',2',3'-tris(acetoxy)heptyl]-2H-py*ran-2-ones (23 and 28).* Treatment of 17 (387 mg, 1.98 mmol) successively with *H<sub>2</sub>O<sub>2</sub>*, OsO<sub>4</sub>, and Ac<sub>2</sub>O according to the previous procedures gave *(via* the saturated lactone 26, and the alcohols 21 and 27, which were not isolated) 23 (62.7 mg, 16%) (identical to that obtained previously) and 28 (198 mg, 51.3%) as a colorless oil. 28: <sup>1</sup>H-NMR: 0.85 <sup>1</sup>*H); 5.03 (ddd, J* = 8, 7, 4, 1 H); *5.30 (ddd, J* = 7, *5, 3,* 2 *H).* IR, MS: identical to those of 23. Anal. calc. for CI8H2,O8: C *58.05,* H 7.58; found: C 57.88, *H* 7.63.  $(t, J = 7, 3H)$ ; 1.2-2.0 (m, 10 H); 2.03 (s, 3 H); 2.12 (s, 3 H); 2.14 (s, 3 H); 2.4-2.7 (m, 2 H); 4.40 (ddd,  $J = 10, 5, 3$ ,

## REFERENCES

- [l] N. C. Franca, J. Polonsky, *C. R. Hebd. Seances Acad. Sci., Ser. C* 1971,273,439.
- [2] **A.** Kjaer, R. Norrestam, J. Polonsky, *Acta Chem. Scand., Ser. B* 1985,39,745.
- [3] M. T. Davies-Coleman, D. E. A. Rivett, *Phytochemistry* 1987,26, 3047.
- [4] L. Van Puyvelde, N. De Kimpe, **S.** Dube, M. Chagon-Dube, Y. Boily, F. Borremans, **F.** Schamp, M. J.O. Anteunis, *Phytochemistry* 1981, 20, 2753.
- [5] J. M. Watt, M. G. Breyer Brandwijk, 'The Medicinal and Poisonous Plants of Southern and Eastern Africa', Livingstone, Edinburgh, 1962, **p.** 516.
- [6] C. Fehr, J. Galindo, *G.* Ohloff, *Helu. Chim. Actu* 1981,64, 1247.
- 171 *C.* W. Jefford, D. Jaggi, J. Boukouvalas, *Tetrahedron Lett.* 1986,27,4011.
- *[8]* **S.** M. Singh, A. C. Oehlschlager, *Can. J. Chern.* 1988,66, 209.
- [9] F. Sato, Y. Kobayashi, 0. Takahashi, T. Chiba, Y. Takeda, M. Kusakabe, *J. Chem. Soc., Chem. Cornmun.*  1985, 1636.
- [lo] F. Sato, 0. Takahashi, T. Kato, Y. Kobayashi, *J. Chem. Soc., Chem. Cornmun.* 1985, 1683.
- [ll] **Y.** Kobayashi, Y. Kitano, T. Matsumoto, F. Sato, *Tetrahedron Lett.* 1986,27,4775.
- [ 121 L. Brandsma, 'Preparative Acetylenic Chemistry', Elsevier Publishing Company, Amsterdam-London-New York, 1971, p.25 and 88.
- [13] F. Sato, *H.* Ishikawa, M. Sato, *Tetrahedron Lett.* 1981,22, *85.*
- [14] F. Sato, *H.* Watanabe, Y. Tanaka, T. Yamaji, M. Sato, *Tetrahedron Lett.* 1983,24, 1041.
- [15] Y. Kobayashi, H. Uchiyama, H. Kanbara, M. Musakabe, F. Sato, *Heterocycles* 1987,25, 549.
- [16] H. Günther, 'NMR Spectroscopy', John Wiley & Sons, New York, 1980.
- [17] K. Mead, T.L. Macdonald, *J. Org. Chem.* 1985, 50, 422.
- [18] 0. Mitsunobu, *Synthesis* 1981, 1.
- [19] K. N. *Houk,* M.N. Paddon-Row, N. Rondan, Y.-D. Wu, F.K. Brown, D. C. Spellmeyer, J.T. Metz, **Y.** Li, R. J. Longcharich, *Science* 1986,231, 1108.
- [20] K. N. Houk, H.-Y. Duh, Y. D. Wu, *S.* R. Moses, *J. Am. Chern. Soc.* 1986,108,2754.
- [21] E. Vedejs,C.K. McClure,J. *Am. Chem. Soc.* 1986,108,1094; E. Vedejs, W.H. Dent,I11, *ibid.* 1989,111,6861.
- [22] J.K. Cha, **W.** J. Christ, Y. Kishi, *Tetrahedron Lett.* 1983, 3943, 3947.
- [23] W. J. Christ, J. K. Cha, Y. Kishi, *Tetrahedron* 1984,40,2247.
- [24] V. Van Rheenen, R. C. Kelly, D. Y. Cha, *Tetrahedron Lett.* 1976,23, 1973.
- [25] M. Schröder, *Chem. Rev.* 1980, 80, 187.
- [26] D. *H.* R. Barton, R. A. *H.* F. Hui, **S.** V. Ley, D. J. Williams, *J. Chem.* Soc., *Perkin Truns. 1* 1982, 1919.
- [27] E. Francotte, D. Lohmann, *Helu. Chirn. Acta* 1987, 70, 1569.